

Dog SAA Sirius SPARCL Kit, Cat. No. SAA-SPL-4

INTRODUCTION

SAA (serum amyloid A) levels increase up to several hundred-fold in dog serum because of injury and infection. It can be used to monitor disease progression. The Sirius SPARCL™¹ assay allows simple and rapid measurement of dog SAA using a Berthold Sirius-L single tube luminometer.

PRINCIPLE OF THE ASSAY

The assay uses two affinity purified dog SAA antibodies. One is conjugated to horseradish peroxidase (HRP), the other to acridan, a chemiluminescent substrate. When HRP and acridan conjugated antibodies bind to SAA they are brought into proximity. Upon addition of hydrogen peroxide, HRP catalyzes oxidation of proximal acridan molecules causing a flash of luminescence that is proportional to SAA concentration.

Because SAA binds to lipoproteins that can interfere with its measurement, serum samples are first mixed with dissociation reagent and incubated for 30-minutes. Dissociated samples are then further diluted. Diluted samples (100 µl) are dispensed into test tubes and mixed with 0.5 ml of combined acridan and HRP conjugates. After 45 minutes, tubes are placed in the Sirius-L luminometer. Luminescence is measured after injection of background reducer, that eliminates nonspecific luminescence, followed by trigger-solution containing hydrogen peroxide. The concentration of SAA is determined from the ratio of sample luminescence to the that of a dog SAA standard.

MATERIALS AND COMPONENTS

Materials provided with the kit:

- Acridan & HRP conjugates, 1 vial **Store at -20°C**
- Dog SAA standard, 1 vial **Store at -20°C**
- Dissociation reagent; DR2-1, 2 ml
- Diluent; CSD50-1, 50 ml
- Background reducer; BR4-1, 4 ml
- Trigger solution; TS4-1, 4 ml
- 15 ml centrifuge tubes, 4

Materials required but not provided:

- Sirius-L luminometer with two injectors and PC
- Precision pipettors and tips
- Test tubes (borosilicate, 12 x 75 mm)
- 12 x 75 mm test tube racks
- Excel software

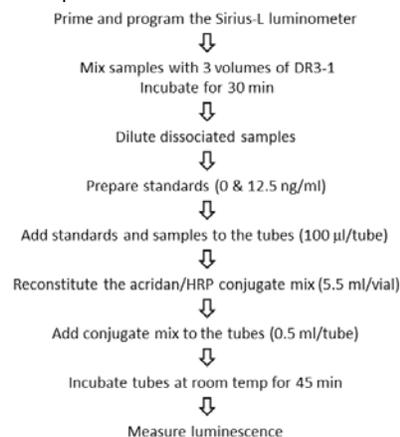
STORAGE

Store the conjugate and standard vials at or below -20°C. The remainder of the kit should be stored at 2-8°C.

GENERAL INSTRUCTIONS

1. Please take the time to completely read all instructions before starting your assay. Contact us if you need clarification.

2. All reagents used in the assay should be removed from the refrigerator or freezer and allowed to reach room temperature (25°C) before use.
3. Follow the sequence of events below when running the assay.



LUMINOMETER SETUP

1. Turn the Sirius-L luminometer on and allow it to warm up for 15 minutes.
2. Place the tubing from injector one into a 15 ml centrifuge tube containing background reducer.
3. Place the tube from injector two into a 15 ml centrifuge tube containing trigger solution.
4. Open the Sirius-L software on the computer. The Protocol Manager screen will load.
5. Click on "Priming Injectors" and program the luminometer to prime injector-1 with 3 x 0.5 ml of background reducer and injector-2 with 3 x 0.5 ml of trigger solution. Make sure to select "to tube" as the destination.
6. Open the luminometer drawer and insert an empty 12 x 75 mm test tube.
7. Close the drawer and click "Prime"
8. Wait for priming to complete then open the drawer and discard the tube.
9. Highlight the Quick Measurement program in the Protocol Manager window and click "Edit". Enter the following parameters:

Delay time:	Measurement time:	Save			
2.6s	0.8s				
Injectors					
#1	Volume [ul]	Delay [s]	#2	Volume [ul]	Delay [s]
<input checked="" type="checkbox"/>	50	1	<input checked="" type="checkbox"/>	100	2

10. Click "Save".
11. Highlight Quick Measurement and click "Run".
12. The luminometer is now ready for use.

¹ SPARCL technology, using acridan- and HRP-conjugated antibodies, was developed by and is licensed from Lumigen Corp.

ASSAY FORMAT

1. Download the Excel spreadsheet from the Life Diagnostics website: <https://lifediagnostics.com/dog-saa-spreadsheet/>
2. The first two rows/tubes are reserved for the zero and 12.5 ng/ml standards.
3. Enter your sample identities and dilution factor(s) into rows 3+ in the order that you intend to measure them.

SAMPLE PREPARATION

Dissociation:

1. Mix 25 μ l of serum with 75 μ l of dissociation reagent DR2-1 (this also provides a 4-fold dilution of the sample).
2. Incubate at room temp for 30 minutes.
3. Use the dissociated samples within 15-min of preparation.

Dilution:

Because serum levels of SAA can range from approximately 0.5 to >100 μ g/ml, we recommend that samples be tested at a minimum of two dilutions. We recommend dilutions of 400 and 16,000-fold.

1. For each sample dispense 495 μ l and 390 μ l of CSD50-1 diluent into separate tubes.
2. Mix 5.0 μ l of dissociated/4-fold diluted serum with 495 μ l of diluent to give a 400-fold dilution.
3. Mix 10.0 μ l of the 400-fold diluted sample with 390 μ l of diluent to give a 16,000-fold dilution.

STANDARD PREPARATION

1. Reconstitute the 12.5 ng/ml SAA standard with the volume of CSD50-1 diluent indicated on the vial label.

CONJUGATE MIX PREPARATION

1. The acridan and HRP conjugate mix should be prepared just before use (step 6 in the Procedure section).
2. Tap the vial to ensure that the contents are at the bottom of the vial before carefully removing the stopper.
3. Add 5.5 ml of diluent CSD50-1 to the vial. Insert the stopper and mix gently by inversion 20x.
4. Each vial of reconstituted conjugate mix provides enough reagent to measure 0 and 12.5 ng/ml standards and up to eight samples.²

PROCEDURE

1. Before starting the assay ensure that the luminometer is primed with background reducer and trigger solution and that the SPARCL program is loaded (refer to the Luminometer Setup section).
2. Prior to starting the assay ensure that the 12 x 75 mm borosilicate test tubes fit easily in the luminometer cuvette holder.
3. Pipet 100 μ l of diluent into tube one. This serves as the zero standard.
4. Pipet 100 μ l of the reconstituted 12.5 ng/ml SAA standard into tube two.
5. Pipet 100 μ l aliquots of the diluted samples into tubes 3, 4, 5... as defined by your assay format.
6. Add 0.50 ml of freshly prepared conjugate mix to each tube and mix gently. A vortex mixer may be used if available.
7. Incubate the mixtures at room temperature.

² If using multiple vials of conjugates to measure more than eight samples, combine the reconstituted contents of all vials and mix briefly before dispensing 0.5 ml

8. After 45 minutes insert tube 1 into the sample holder of the luminometer and close the drawer. The luminometer automatically injects background reducer and trigger solution, then measures luminescence (RLU/s).
9. Once the RLU/s value is recorded on the screen open the drawer and discard the tube.
10. Determine luminescence for the remaining tubes.

CALCULATION OF RESULTS

1. Cut and paste the RLU/s values into the RLU/s column of the downloaded Excel spreadsheet.
2. SAA values (μ g/ml) are automatically calculated.

TYPICAL RESULTS

Typical results for the 0 and 12.5 ng/ml standards and four dog serum samples are shown below.

Dog SAA SIRIUS Worksheet						
Tube	Sample ID	Comments	Dilution	RLU/s	Zero subtracted RLU/s	μ g/ml
1	0 ng/ml SAA			12840	0	
2	12.5 ng/ml SAA			134430	121590	
3	1		400	102992	90152	3.71
4	2		400	96316	83476	3.43
5	3		400	34045	21205	0.87
6	4		16000	59857	47017	77.34

LUMINOMETER MAINTENANCE

The luminometer injectors should be cleaned with distilled or deionized water at the end of each day of use.

1. Exit the Quick Measurement screen and click on "Priming Injectors". Enter the following parameters:

2. Click "Prime". The contents of the injectors will be returned to the respective tubes. If future use is intended store the sealed tubes in a refrigerator.
3. Place the tubing from injectors one and two into separate tubes containing distilled or deionized water.
4. Enter the following parameters:

5. Insert an empty 12 x 75 mm test tube into the luminometer. Close the drawer and click "Prime".
6. After priming is complete discard the 12 x 75 mm tube and its contents.

aliquots into the reaction tubes. Larger volumes of background reducer and trigger solution must also be used.